

# The safety of a therapeutic product composed of a combination of stem cell released molecules from adipose mesenchymal stem cells and fibroblasts

Greg Maguire\*, 1 Peter Friedman 1,2

- <sup>1</sup>Bio Regenerative Sciences, Inc. San Diego, CA 92121, USA
- <sup>2</sup>Animal BioSciences, LLC Bartow, FL 33830, USA
- \*Author for correspondence: gmaguire@bioregenerativesciences.com

Aim: We sought to determine the safety profile of a therapeutic candidate composed of the released molecules from a combination of human adipose-derived mesenchymal stem cells and fibroblasts. Although stem cells, their progenitor cells and the molecules that are released from these cells have some demonstrated therapeutic value, much more needs to learn about the efficacy, mechanism of action and the safety profiles of these stem cell-based therapeutics. Methods: A number of cellular, in vitro, in vivo and human studies were performed to analyze cellular, tissue and systemic safety profiles of the combinatorial product. Results: At the levels tested in this study, ranging from demonstrated therapeutic doses to supratherapeutic doses, the combinatorial product demonstrated an excellent safety profile in all in vitro, cellular, tissue and systemic studies. Conclusions: We found evidence that a therapeutic candidate composed of the molecules released from human adipose-derived mesenchymal stem cells and human fibroblasts has an excellent safety profile, and that the product warrants further studies for safety and efficacy where dosing may include topical application, injection and oral application.

Lay abstract: Stem cell transplants have demonstrated life-saving capabilities for some diseases, and the molecules released from stem cells are currently in therapeutic development for a number of conditions. Stem cell science is a relatively new science and is in need of a better understanding of mechanisms of action and acute and long-term safety profiles. Here, we performed a number of safety tests for stem cell released molecules from a combination of adipose-derived mesenchymal stem cells and fibroblasts that have demonstrated efficacy in a number of conditions. Using *in vitro*, *in vivo* and skin sensitivity studies in humans, the stem cell therapeutic was found to have an excellent safety profile when tested for toxicity, mutagenicity, tumorigenesis, ocular toxicity, inflammation and irritation.

First draft submitted: 20 February 2020; Accepted for publication: 20 April 2020; Published online: 29 May 2020

Keywords: inflammation • stem cells • stem cell safety • stem cell secretome • toxicity • tumorigenesis

Stem cell transplants have been used for over 40 years with demonstrated life-saving capabilities for some blood diseases [1], and the molecules and exosomes released from the stem cells are currently in therapeutic development for a number of diseases and conditions, including neurodegenerative diseases [2], heart conditions [3], glaucoma [4], hearing loss [5] and skin diseases [6,7]. However, stem cell science is a relatively new science, and therapeutic development using stem cells, even approved stem cell therapies for blood diseases, is in need of a better understanding of mechanisms of action and acute and long-term safety profiles, both for the cells and their released molecules. Approved bone marrow stem cell (BMSC) transplants have many associated risks, including possible induction of cancer [8], including skin cancer [9], the transference of cancer cells from the donor to the patient [10] – given the bone marrow is a site of recirculated cancer cells [11,12] – transference and engraftment of BMSCs with pathogenic mutations [13], and aging of the tissue in which the implant occurs [14]. Many factors, which are often overlooked, must be considered when developing stem cell-based therapeutics, including something as fundamental as the



choice of stem cell type where adipose mesenchymal stem cells (ADSCs) have many advantages over BMSCs for therapeutic development [15]. For example, even the molecules from BMSCs may induce cancer [16], whereas from ADSCs do not. Moreover, ADSCs can be used for expressing and delivering targeted therapeutics [17], including the treatment of cancer [18], for example, lung cancer and melanoma [19], osteoarthritis [20] and cardiovascular disease [21].

Here, we performed a number of safety tests for a stem cell-based therapeutic comprised of the stem cell released molecules (secretome) from a combination of ADSCs and fibroblasts (FBs), the combination of which has demonstrated efficacy in a number of conditions [2,7] and is conceptually based on developing a system therapeutic [22] for the physiological renormalization of tissue in various disease states or abnormal conditions [23]. Using *in vitro*, *in vivo* and skin sensitivity studies in humans, the stem cell therapeutic comprised of stem cell released molecules from ADSCs and FBs was determined to have an excellent safety profile when tested for *in vitro* and *in vivo* toxicity, the Ames mutagenicity assay, *in vivo* tumorigenesis, *in vivo* inflammation, ocular histology and a human skin patch test for irritation and allergic reaction.

#### Method

Because a combination of the secretome from ADSCs and FBs has been shown to be efficacious for several conditions [2,7], we performed a number of safety studies for the combination to help clarify its use in humans under acute and chronic conditions.

Stem cell culture procedures have been described previously [2] and performed using the basic procedures described in the patent numbers 9545370 and 9446075. We used a 50–50% mix of the molecules released from the two cell types. All processes were performed using GMP procedures. Briefly, a proprietary collection of stem cell lines derived from human skin were cultured using no penicillin/streptomycin, and 1% fetal bovine serum, under hypoxic conditions. When cultures reached confluence, they were passaged for a limited number of times (fewer than 10) before disuse. Total conditioned medium from the multiple cell types, containing a soluble fraction and an exosome fraction, was harvested at each passage and the passages combined into one batch for product development. Parts of our stem cell technology used here are covered by US patents [46–49].

# Safety studies, human repeat insult patch test & canine repeat insult patch test

The canine repeat insult patch test and human repeat insult patch test [24] were used to assess primary and accumulative irritation, and/or allergic contact sensitization, in both male and female human subjects between the ages of 18 and 66. All subjects were free of skin disease and were prohibited from using topical or systemic antihistamines and steroids, beginning 7 days prior to the onset and throughout the duration of the study. Measurements were performed by a trained rater technician. For the induction phase of the study, the patches. 0.1 g of the combinatorial secretome per square inch Webril dressing, were applied three-times weekly for 3 weeks, for a total of nine applications. The patches were removed 48 h after application and evaluated before a fresh patch was reapplied to the same area. For the challenge phase of the study, 2 weeks after the final induction phase patch was applied, an adjacent, virgin area of skin then received a fresh patch and was evaluated at 24 and 72 h following application of the patch. Ratings were scored using the following table:

- 0 No visible skin reaction.
- 1 Barely perceptible or spotty erythema.
- 2 Mild erythema covering most of the test site.
- 3 Moderate erythema, possible mild edema.
- 4 Marked erythema, possible edema.
- 5 Severe erythema, possible edema, vesiculation, bullae and/or ulceration.

For the canine repeat insult patch test study, 27 qualified subjects absent of skin disease or irritation and no use of medications, male and female, ranging from age 2 to 7 years were studied. Stem cell released molecules from the ADSCs and FBs (S2RM) was applied to the belly of the dog, held for 2 min to allow the S2RM to absorb into the skin, and no patch applied. The rater was a practicing veterinarian, who used the same rater scale that was used in the HIRPT. Photos were taken following each application, which was performed three-times weekly for 3 weeks. All 27 dogs finished the study.

# **Oral toxicity studies**

This toxicity study used three groups of 3 male and 3 female Sprague–Dawley rats (18 total). Once-daily oral (gavage) administration was as follows: Group 1 was administered the vehicle only once daily at 4.4 ml/kg/dose for 28 consecutive days. Group 2 was administered S2RM once daily at 0.44 ml/kg/dose for 28 consecutive days. Group 3 was administered S2RM once daily at 4.4 ml/kg/dose for 28 consecutive days. As a biomarker for a possible inflammatory response to the S2RM administration, all rats were subjected to blood collection (Red-Top) for ELISA testing for the pro-inflammatory cytokines IL-10 and IL-31, which measured prior to the first dose (Day 1) and on the day following the last dose (Day 29).

#### **Results**

## Batch reproducibility: total protein

A shotgun approach to a proteomic analysis has yet to be performed, but in 10 samples from various batches, we measured, using a microarray, 18 proteins in an exosome fraction present in each. The presence of the 18 proteins and exosomes in each batch, along with strict standard operating procedures (SOPs) and low variability in total protein count from batch to batch suggests that each batch is similar. RNA and metabolites have not yet been measured. The Bradford method was used in all protein determinations. The Bradford protein assay is a spectroscopic analytical procedure used to measure the concentration of protein in a solution. Samples, performed in duplicate, from a total of 15 batches of S2RM were analyzed. The variation between duplicates in all cases was less than 10%. Five samples of different S2RM batches and five samples of the individual batches of SRM from the individual cell lines that make-up the S2RM were analyzed for total protein. All samples were taken from frozen aliquots of batches previously used to make skincare products with demonstrated efficacy. All batches produced at least 500 ug/ml of total protein, with a high value in one batch of 630 ug/ml. The mean value of all batches was 554 ug/ml. The variability in all the batches was 20% or less. With the exception of one batch displaying a high protein count (630 ug/ml), the rest of the batches had a variability of 12% or less.

### Skin safety testing, irritation

Totally, 91 participants, both male and female and ranging in age from 18 to 66 years, were selected for this evaluation. A total of 50 participants completed the study. The remaining subjects discontinued their participation for various reasons, none of which were related to the application of the test material. Of the 50 subjects completing the study, all were rated at 0 during both the induction phase and the challenge phase, indicating that the S2RM induced no immediate or long-term irritation, or allergic reaction. Because the S2RM is also intended for therapeutic development to treat companion animals, we tested for cross-species irritation in canines. Similar to that for humans, no irritation or allergic reaction was observed in the 27 canines, as all scored '0' throughout the testing period.

# Analysis of ocular pathology & tumorigeneicity following topical application of S2RM to the eye

Both eyes were recovered from all animals following seven consecutive days of twice daily administration of the S2RM as eye drops applied to the cornea. Eyes were submitted to Colorado Histo-Prep for histopathological evaluation by a board-certified veterinary pathologist who had no knowledge of which eyes had been treated. Six rabbits (New Zealand White) received the S2RM solution in one eye, and the other eye served as a control. One control rabbit receiving no S2RM in either eye was also was also used. The samples were trimmed, processed, embedded, sectioned and stained. Histopathology of the tissues was conducted on slides stained with hematoxylin and eosin with an emphasis on eye irritation and inflammation.

### Oral administration toxicity, tumorigenesis & inflammation results

No significant differences in organ weights were found in the male or female rats when compared with controls (Figure 1A & B). When compared with the control group, there were no biologically relevant changes from Day 1 to Day 29 for group average IL-10 or IL-31 in either SRM-treated group, suggesting that the S2RM did not induce an inflammatory reaction when administered orally. All tissues were also analyzed for tumor formation, and no tumors were found in any of the analyzed tissues (analyzed tissues are depicted in Figure 1). As a general indicator of health, bodyweights were recorded prior to the first dose and weekly thereafter, including the day of euthanasia (Figure 1C). When compared with the control group, there was mildly decreased bodyweight gain in high-dose (SRM at 4.4 ml/kg/dose) males and females over the course of the study. Bodyweight in the S2RM

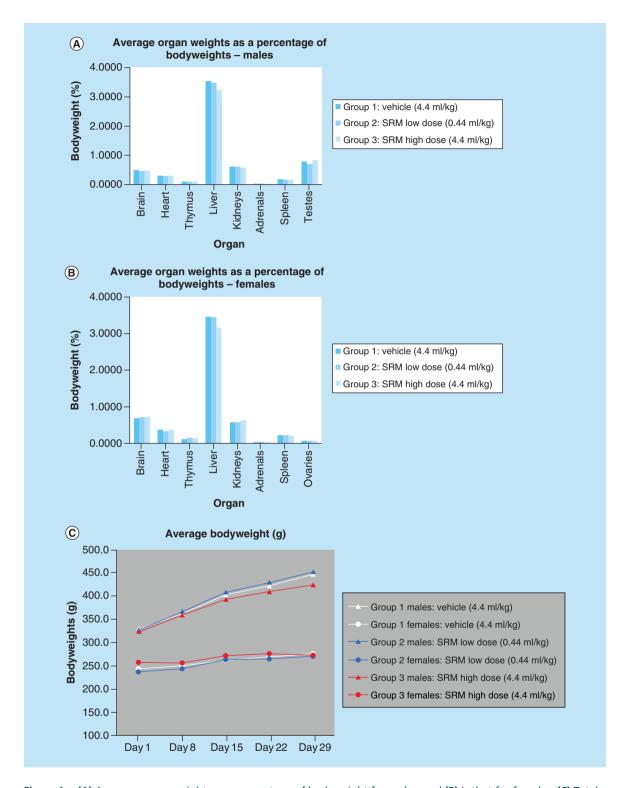


Figure 1. (A) Average organ weights as a percentage of bodyweight for males, and (B) is that for females. (C) Total bodyweight over a 30-day period during dosing by oral gavage of S2RM. No significant differences in weights were observed for any of the organs, and none of the organs showed evidence of toxicity or tumorigenesis SRM: Stem cell released molecules; S2RM: Stem cell released molecules from the adipose mesenchymal stem cells and fibroblasts.

A								
Sector	Characteristics (expected)	TA97a	TA98	TA100	TA1535			
I	Histidine (no growth)	NG	NG	NG	NG			
II	rfa mutation CV (zonal inhibition)	S	S	S	S			
III	+AMP	G	G	G	G			
IV	+AMP and +TET	NG	NG	NG	NG			
N/A	uvrB/uvrA-UV light	NG	NG	NG	NG			
N/A	Titer (organisms/50 μl)	15	59	84	10			
В								
		Salmonella typhimurium						
	TA97a		TA98		TA100		TA1535	
	CFP	Mean	CFP	Mean	CFP	Mean	CFP	Mean
Negative control without S9	64 51	58	25 26	26	24 52	28	24 21	23
Negative control with S9	65	71	31 34	33	33 26	30	18 18	18
ICR 191 acridine without S9 positive control	163 203	180						
Daunomycin without S9 positive control			156 165	160				
Sodium azide without S9 positive control					67 74	71	44 72	58
Mitomycin C without S9 positive control								
2-aminoanthracene with S9 positive control	358 342	350	430 404	420	155 127	140	93 92	93
Product solution without S9 concentration of 5 mg/plate	85 99 94	93	24 16 18	19	25 31 35	30	17 18 15	17
Product solution with S9 concentration of 5 mg/plate	119 85 103	100	19 40 39	33	26 51 35	37	24 31 21	25

A twofold or greater increase in the number of mean revertants for the S2RM over the mean number of revertants obtained from the negative controls was considered mutagenic. (A) Strain characteristics and standard strain plate counts. (B) Standard plate incorporation assay – reversion rates for tester strains.

S2RM: Stem cell released molecules from the adipose mesenchymal stem cells and fibroblasts.

Blank cells indicate 'not applicable.

CFP: Counts from plate; G: Growth; Mean: Average of plate; NG: No growth; S: Sensitive; S: Slight growth; S2RM: Stem cell released molecules from the ADSCs and FBs.

low-dose group (Group 2, 0.44 ml/kg/dose) was comparable to the control group throughout the study in males and females. Therefore, the general health and the appetite of the experimental animals were normal.

# Results of ocular pathology & tumorigeneicity following topical application of S2RM to the eye

No S2RM-related lesions or tumors were observed in this study after 7 days of topical S2RM/placebo treatment and prior to euthanasia. The eye globe sections were of very good quality and no differences in cornea morphology were detected in any specimen. Overall eye structure was normal, and no ocular opacities were detected.

# Ames test

The Ames reverse mutation was used in four strains of *Salmonella*. Mutagens identified using the Ames test are usually carcinogens, given Ames showed an association of carcinogenicity and mutagenicity of 90%; in other words, most carcinogens are also mutagenic (Table 1) [25]. In the four strains of bacteria tested (Figure 1), the S2RM was shown to be nonmutagenic tested with or without S9 activation. S9 is a crude liver enzyme extract that was used to convert materials without any genotoxic activity to active genotoxic entities; in other words, to duplicate the possible liver derived metabolites of the test product that are mutagenic.

Table 2. Using a mouse cell line L929, the S2RM conditioned media is shown to have no direct cytotoxic effects on the cells (fibroblasts).

Sample	Malformation	Degeneration	Sloughing	Lysis	Reduction in cell layer density
Sample – 24 h	0.0	0.0	0.0	0.0	0.0
48 h	0.0	0.0	0.0	0.0	0.0
Negative control – 24 h	0.0	0.0	0.0	0.0	0.0
48 h	0.0	0.0	0.0	0.0	0.0
Media control – 24 h	0.0	0.0	0.0	0.0	0.0
48 h	0.0	0.0	0.0	0.0	0.0
Positive control – 24 h	2.2	2.2	2.2	2.2	2.4
48 h	4.4	4.4	4.4	4.4	4.4

Evaluation of results: after incubation, the biological reactivity (cellular degeneration and malformation) is described and rated on a scale of 0–4 as follows: grade reactivity description of reactivity zone. 0, none (no detectable zone under or around specimen); 1, slight (some malformed or degenerated cells under specimen); 2, mild (zone limited to area under specimen); 3, moderate (zone extends 0.5 to 1.0 cm beyond specimen); 4, severe (zone extends greater than 1.0 cm beyond specimen but does not involve the entire dish). If both test articles exhibit reactivity grades of 0, 1 or 2, the sample is noncytotoxic. If both test articles exhibit reactivity grades of 3 or 4, the sample is cytotoxic. If only one (1) test articles exhibits a grade 3 or 4, the test may be repeated with four (4) test articles. To be noncytotoxic, none of the four retest articles may exhibit a grade of 3 or 4.

SZRM: Stem cell released molecules from the adipose mesenchymal stem cells and fibroblasts.

Cample		IDE Score	gen, keratin and other dermal proteins.		
Sample	Dose		Predicted ocularIrritancy classification		
S2RM	25 μΙ	9.6 <sup>A</sup>	Minimal irritant		
S2RM	50 μl	7.1	Minimal irritant		
S2RM	<b>75</b> μl	5.5	Minimal irritant		
S2RM	100 μΙ	4.8	Minimal irritant		
S2RM	125 μl	3.0	Minimal irritant		
A. Maximum qQualified score	2				
IDE	Predicted ocular irritancy	Predicted ocular irritancy classification			
0.0-12.5	Minimal irritant	Minimal irritant			
12.5–30.0	Mild irritant				
30.0-51.0	Moderate irritant				
51.0-80.0	Severe irritant				

## Cellular cytotoxicity in a mouse L929 FB cell line

FBs exist in most tissue compartments throughout the body, particularly in the skin where the current S2RM technology has proven to be efficacious in a number of skin conditions [7]. We, therefore, tested the S2RM for potential cytotoxic effects in FBs. Table 2 demonstrated the data providing evidence that the S2RM has no cytotoxic effects in FBs as measured by observing changes in cell structure or the number of cells in the culture.

# Protein misfolding tests (Dermal Irritection® Assays System)

In this study, the Irritection<sup>®</sup> system from InVitro International (CA, USA) was used for the detection of misfolding and structural changes in proteins. Using this methodology, an irritant chemical will disrupt the ordered structure of keratin and collagen and result in the release of a bound indicator dye. Additionally, irritants will induce changes in the conformation of the globular proteins found in the reagent solution. The extent of dye release and protein denaturation was quantitated by measuring the changes in the optical density of the S2RM solution at 450 nm (OD450). The results provide evidence that S2RM induced no structural changes or misfolding of the test proteins (Table 3).

## Heavy metals

Numerous products and procedures are involved in the processing of the stem cells for the collection of their molecules, presenting a number of points were contamination may take place. We, therefore, tested for the presence



Table 4. Testing for the presence of four	common heavy metals using inductively coupled plasma mass spectrometry.
Test	Result
Arsenic	<0.025 p.p.m.
Cadmium	<0.025 p.p.m.
Lead	<0.025 p.p.m.
Mercury	<0.025 p.p.m.
None of the heavy metals tested were present in significant, up.p.m.: Part per million.	nsafe amounts.

of four common heavy metals in the S2RM end-product used as the therapeutic. No significant level of heavy metals was present in the S2RM (Table 4).

#### Discussion

The results presented here for a combination therapeutic containing the stem cell-released molecules from ADSCs and FBs provide evidence that these molecules are safe whether they are administered orally or topically to the eye or skin. No signs of toxicity, tumorigenesis, mutagenicity, irritation and allergic reaction or inflammation were measured when the S2RM was compared with controls. These data are also important for stem cell transplants because when transplanted stem cells graft in the tissue and remain alive and healthy for a long time [26], much of the cell's therapeutic effect is mediated by the release of molecules that act as paracrine and autocrine factors [27,28]. Understanding the cellular effects versus the paracrine effects of stem cell therapy is important for the development of therapeutics, both in terms of efficacy as well as safety. For example, we know that BMSC transplants induce aging of the implanted tissue [14], and increase the probability of cancer [8,29], but we do not understand the mechanisms by which this happens. Is it the choice of the BMSC type versus the ADSC type that is responsible for these issues [30], or the implantation process of exogenous cells, or the release of specific molecules from the BMSCs that cause these adverse events? Although we have no evidence to answer the aforementioned questions directly, in this study, we do provide an evidence that the molecules from ADSCs and FBs are safe for therapeutic development. To help answer these questions, other studies have shown that the secretome from BMSCs induces tumorigenesis in an in vivo and in vitro mouse model through activation of mTOR [31], whereas ADSCs and their conditioned media inhibit cancer growth using in vivo and in vitro mouse models [32], suggesting that ADSCs are a better cell choice for therapeutic development than are BMSCs in terms of tumorigenesis. ADSCs also present numerous other benefits compared with BMSCs for a variety of safety and efficacy issues [15]. However, more data are needed to understand the long-term consequences of the S2RM on various health parameters, and whether injection and intravenous (IV) administration will present a like safety profile as observed in the current studies.

In the development of stem cell therapeutics, Goldring et al. [33] asked the question of whether we are setting a higher bar for the clinical implementation of stem cell-derived therapeutics than we currently apply for other types of cellular therapy. The authors argue there is a danger that if perfection is a prerequisite for beginning stem cell therapeutics, then we will never begin. As Maguire [30] has argued, if the current hype about stem cell therapies continues, where unapproved therapies without a knowledge of risk or reward are burgeoning [34], without a knowledge of the procedure's risks, then a proper evaluation of the risk versus reward ratio cannot be made for that particular stem cell therapy. Regardless of where the bar is set for other therapies, where medical procedures, like most drugs, have unknown or hidden long-term consequences to health, the risks must be evaluated as best as possible so informed risk versus reward decisions can be made. Often, when considering marketed drugs, not until Phase IV postmarket approval, are the long-term consequences of a drug discovered. This is exemplified by the many drugs pulled from market or with safety issues 3-4 years after their approval [35,36]. Even more unfortunate, the problem is worse with medical procedures [37]. Such is the case with approved stem cell transplants. Many case studies have reported the approved stem cell transplants to be associated with the later development of cancer [8], and unapproved stem cell transplant procedures are notorious for adverse side-effects, including development of cancer [38]. The effects of approved BMSC transplant in cancer relapse are not well understood but are thought to involve epigenetic factors in the stem cells used for the transplant [29]. In addition, BMSC transplants may cause aging of the tissue as measured in T cells using a p16 biomarker [14], indicating the increased level of cellular senescence in the surrounding tissue, a factor in the increased probability of tumorigenesis [39].

Although stem cell therapy is in a period of rapid advancement, the science of stem cell safety assessment must also evolve, not to hinder progress, rather to support, guide and expedite the progress of patient treatment using stem cell-based technologies. The development of a rich safety database is necessary to ensure that we can proceed with appropriate safeguards in place and allow that stem cell-based therapeutic approaches develop in a way that benefits society overall by using well supported, data-driven risk versus reward analysis. These data presented here are one such needed set of safety data to evaluate the patient risk versus benefit ratio for stem cell therapeutics in general, and specifically for our combination of stem cell released molecules from ADSCs and FBs.

# **Future perspective**

To date, stem cell therapeutics have heavily focused on using stem cell transplants. However, for many conditions where stem cells have been used as the therapeutic, most of the therapeutic benefit of the stem cell transplant was because of the many molecules released by the transplanted stem cells. Many medical procedures have little or no reward, and only present risk to the patient [37]. Indeed, many stem cell transplants procedures have inherent negative consequences to the patient, including a number of safety issues.

The molecules and exosomes released from certain stem cell types will provide a more efficacious, safe, less expensive and easier to dose therapeutic than do stem cells for a number of conditions. In the next decade, we are likely to optimize the methods to store and process the exosomes from stem cells, such as freezing methods [40] and lyophilization [41,42]. This knowledge will help to facilitate the formulation, transportation, and 'off the shelf' use of exosomes for therapeutic application, obviating many of the difficulties associated with cellular therapies [43]. Given that molecules released from specific stem cell types, such as BMSCs and ADSCs, and their various phenotypes, can preferentially elicit inflammatory, infection fighting pathways (BMSCs) or proresolving, wound healing pathways (ADSCs) [15], the choice of stem cell type and its specific phenotype will be critical to their therapeutic development for particular diseases and conditions. For example, in many viral diseases, including COVID-19, the disease is initially characterized by high viral loads in tissues, such as pulmonary and cardiovascular tissues, followed by clearance of the virus. This can lead to sequelae after the viral clearance, due a dysregulated innate and adaptive immune response, which can include a life threatening 'cytokine storm.' Given that adult stem cells are part of and regulate the innate and adaptive immune systems [44], and that some stem cell types are better at fighting infection, while other types are better at resolving inflammation and repairing tissue, a therapeutic regimen for COVID-19 may include an early phase dosing of exosomes from BMSCs to fight infection [15], and a later phase dosing of exosomes from ADSCs once the virus has cleared to resolve inflammation and rebuild the damaged tissue, with the potential to reduce fibrosis [15] in the alveoli and heart tissue [45].

Given there are currently no approved exosome therapies, future work will need to test these ideas in animal models that best predict outcomes in humans, and to strategize with the Federal Drug Administration, EMA and other regulatory bodies to develop the best chemistry, manufacturing and controls methods and clinical tests to bring these stem cell released molecules/exosomes to the market as approved therapeutics for defined health indications.

# **Summary points**

- A combination of the molecules from adipose mesenchymal stem cells and fibroblasts were tested for their safety
  as a therapeutic.
- A Repeat Insult Patch Test in humans demonstrated no irritation.
- No ocular pathology or tumorigeneicity was observed.
- Oral administration resulted in no tumorgeneisis, toxicity, or inflammation.
- The Ames test for mutagenicity was negative.
- There was no cellular toxicity in a fibroblast cell line.
- No misfolding or structural changes in collagen and keratin proteins was measured.
- Mass spectrometry analysis measured no significant amount of heavy metals.

#### **Author contributions**

G Maguire wrote this paper and helped design and perform the experiments. P Friedman helped design and perform the experiments.

#### Acknowledgments

The authors thank L Green for her expert help in cell culture.

#### Financial & competing interests disclosure

G Maguire has equity in BioRegenerative Sciences, Inc. P Friedman has equity in Animal BioSciences and BioRegenerative Sciences Inc. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

#### Ethical conduct of research

The study design and animal usage were reviewed and approved by the CARE Research Institutional Animal Care and Use Committee (IACUC) for compliance with regulations prior to study initiation (IACUC Number 1730). Animal welfare for this study was in compliance with the US Department of Agriculture's (USDA) Animal Welfare Act (9 CFR Parts 1, 2 and 3), the Guide for the Care and Use of Laboratory Animals, [1] and CARE Research SOPs. The authors state that they have obtained appropriate institutional review board approval or have followed the principles outlined in the Declaration of Helsinki for all human or animal experimental investigations. In addition, for investigations involving human subjects, informed consent has been obtained from the participants involved.

The authors state that they have obtained verbal and written informed consent from the patient/patients for the inclusion of their medical and treatment history within this case report.

#### Open access

This work is licensed under the Creative Commons Attribution 4.0 License. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/

### References

- Thomas ED, Lochte HL, Cannon JE, Sahler OD, Ferrebee JW. Supralethal whole body irradiation and isologous marrow transplantation in man. J. Clin. Invest. 3, 1709–1716 (1959).
- Maguire G, Paler L, Green L et al. Rescue of degenerating neurons and cells by stem cell released molecules: using a physiological renormalization strategy. Physiol. Rep. 7(9), e14072 (2019).
- 3. Marban E. A mechanistic roadmap for the clinical application of cardiac cell therapies. Nat. Biomed. Eng. 2(6), 353–361 (2018).
- 4. Klingborn M, Dismuke WM, Bowes Rickman C, Stamer WD. Roles of exosomes in the normal and diseased eye. *Prog. Retin. Eye Res.* 59, 158–177 (2017).
- 5. Qui Y, Qui J. Stem cells: a new hope for hearing loss therapy. Adv. Exp. Med. Biol. 1130, 165–180 (2019).
- Wang L, Hu L, Zhou X et al. Exosomes secreted by human adipose mesenchymal stem cells promote scarless cutaneous repair by regulating extracellular matrix remodeling. Sci. Rep. 7, Article Number: 13321 (2017).
- Maguire G, McGee Paler S et al. Homeostatic renormalization of skin in traumatic, irradiation, autoimmune, and aging conditions using S2RM stem cell released molecules enhances healing and reduces pain. International J Medical Res. (2020) (In Press).
- 8. Cooley LD, Sears DA, Udden MM *et al.* Donor cell leukemia: report of a case occurring 11 years after allogeneic bone marrow transplantation and review of the literature. *Am. J. Hematol.* 63(1), 46–53 (2000).
- Omland SH, Gniadecki R, Hædersdal M, Helweg-Larsen J, Omland LH. Skin cancer risk in hematopoietic stem-cell transplant recipients compared with background population and renal transplant recipients. A population-based cohort study. *JAMA Dermatol*. 152(2), 177–183 (2016).
- Araf S, Wang J, Margaret Ashton-Key M et al. Transmission of diffuse large B-cell lymphoma by an allogeneic stem-cell transplant. Haematologica 104, e174–e177 (2019).
- 11. Ishikawa F, Yoshida S, Saito Y et al. Chemotherapy-resistant human AML stem cells home to and engraft within the bone-marrow endosteal region. Nat. Biotechnol. 25(11), 1315–1321 (2007).
- 12. Roato I, Ferracini R. Cancer stem cells, bone and tumor microenvironment: key players in bone metastases. *Cancers (Basel)* 10(2), E56 (2018).
- 13. Wong WH, Bhatt S, Trinkaus K et al. Engraftment of rare, pathogenic donor hematopoietic mutations in unrelated hematopoietic stem cell transplantation. Sci. Transl. Med. 12(526), eaax6249 (2020).
- 14. Wood WA, Krishnamurthy J, Mitin N et al. Chemotherapy and stem cell transplantation increase p16<sup>INK4a</sup> expression, a biomarker of T-cell aging. EBioMedicine 11, 227–238 (2016).
- 15. Maguire G. The safe and efficacious use of secretome from fibroblasts and adipose-, but not bone marrow-, derived mesenchymal stem cells for skin therapeutics. *J. Clin. Aesthet. Dermatol.* 12(8), E57–E69 (2019).

#### Research Article Maguire & Friedman

- Liu C, Feng X, Wang B et al. Bone marrow mesenchymal stem cells promote head and neck cancer progression through periostin-mediated phosphoinositide 3-kinase/Akt/mammalian target of rapamycin. Cancer Sci. 109(3), 688–698 (2018).
- 17. Mirzeai H, Sahebkar A, Avan A et al. Application of mesenchymal stem cells in melanoma: a potential therapeutic strategy for delivery of targeted agents. Curr. Med. Chem. 23(5), 455–463 (2016).
- 18. Mohammadi M, Jaafari MR, Mirzaei HR, Mirzaei H. Mesenchymal stem cell: a new horizon in cancer gene therapy. *Cancer Gene Ther.* 23(9), 285–286 (2016).
- 19. Mirzeai H, Salehi H, Oskuee RK *et al.* The therapeutic potential of human adipose-derived mesenchymal stem cells producing CXCL10 in a mouse melanoma lung metastasis model. *Cancer Lett.* 419, 30–39 (2018).
- 20. Mianehsaz E, Mirzaei HR, Mahjoubin-Tehran M et al. Mesenchymal stem cell-derived exosomes: a new therapeutic approach to osteoarthritis? Stem Cell Res. Ther. 10(1), 340 (2019).
- 21. Goradel NH, Hour FG, Negahdari B *et al.* Stem cell therapy: a new therapeutic option for cardiovascular diseases. *J. Cellular Biochem.* 119, 95–104 (2018).
- 22. Maguire G. Systems biology approach to developing "systems therapeutics". ACS Med. Chem. Lett. 5(5), 453-455 (2014).
- 23. Maguire G. Physiological renormalization using systems therapeutics. Future Sci. OA 6(1), FSO428 (2020).
- 24. McNamee PM, Api AM, Basketter DA et al. A review of critical factors in the conduct and interpretation of the human repeat insult patch test. Regul. Toxicol. Pharmacol. 52(1), 24–34 (2008).
- 25. McCann J, Choi E, Yamasaki E, Ames BN. Detection of carcinogens as mutagens in the Salmonella/microsome test: assay of 300 chemicals. *Proc. Natl Acad. Sci. USA* 72(12), 5135–5139 (1975).
- Scala S, Basso-Ricci L, Dionisio F et al. Dynamics of genetically engineered hematopoietic stem and progenitor cells after autologous transplantation in humans. Nat. Med. 24, 1683–1690 (2018).
- 27. Chimenti I, Smith RR, Li TS et al. Relative roles of direct regeneration versus paracrine effects of human cardiosphere-derived cells transplanted into infarcted mice. Circ. Res. 106, 971–980 (2010).
- 28. Riazifar M, Pone EJ, Lötvall J, Zhao W. Stem cell extracellular vesicles: extended messages of regeneration. *Ann. Rev. Pharmacol. Toxicol.* 57, 125–154 (2017).
- 29. Christopher MJ, Petti AA, Rettig MP et al. Immune escape of relapsed AML cells after allogeneic transplantation. N. Engl. J. Med doi: 10.1056/NEJMoa1808777 (2018) (Epub ahead of print).
- 30. Maguire G. Transplanted stem cells survive a long time do they make you sick? J. R. Soc. Med. 112(10), 412-414 (2019).
- Liu C, Feng X, Wang B et al. Bone marrow mesenchymal stem cells promote head and neck cancer progression through Periostin-mediated phosphoinositide 3-kinase/Akt/mammalian target of rapamycin. Cancer Sci. 109(3), 688–698 (2018).
- 32. Cousin B, Ravet E, Poglio S et al. Adult stromal cells derived from human adipose tissue provoke pancreatic cancer cell death both in vitro and in vivo. PLoS ONE 4, e6278 (2009).
- 33. Goldring CEP, Duffy PA, Benvenisty N et al. Assessing the safety of stem cell therapeutics. Cell Stem Cell 8, 618-628 (2011).
- 34. Turner L, Knoepfler P. Selling stem cells in the USA: assessing the direct-to-consumer industry. Cell Stem Cell 19, 154-157, (2016).
- ProCon. 35 FDA-approved prescription drugs later pulled from the market. ProCon, (2014). https://prescriptiondrugs.procon.org/fda-approved-prescription-drugs-later-pulled-from-the-market/
- 36. Downing NS, Shah ND, Aminawung JA *et al.* Postmarket safety events among novel therapeutics approved by the US Food and Drug Administration between 2001 and 2010. *JAMA* 1854–1863 doi:10.1001/jama.2017.5150 (2017) (Epub ahead of print).
- 37. Kumar S, Nash DB Health care myth busters: is there a high degree of scientific certainty in modern medicine? Sci. Am. (2011).
- 38. Diouhy BJ, Awe O, Rao RC, Kirby PA, Hitchon PW. Autograft-derived spinal cord mass following olfactory mucosal cell transplantation in a spinal cord injury patient. *J. Neurosurg. Spine* DOI: https://doi.org/10.3171/2014.5.SPINE13992 (2014) (Epub ahead of print).
- 39. Schosserer M, Grillari J, Breitenbach M. The dual role of cellular senescence in developing tumors and their response to cancer therapy. Front. Oncol. 7, 278 (2017).
- 40. Jeyaram A, Jay SM Preservation and storage stability of extracellular vesicles for therapeutic applications. AAPS J. 20(1), 1 (2017).
- Charoenviriyakul C, Takahashi Y, Nishikawa M, Takakura Y Preservation of exosomes at room temperature using lyophilization. Int. J. Pharm. 553(1–2), 1–7 (2018).
- 42. Bari E, Perteghella S, Catenacci L et al. Freeze-dried and GMP-compliant pharmaceuticals containing exosomes for acellular mesenchymal stromal cell immunomodulant therapy. Nanomedicine 14(6),753–763 (2019).
- 43. Maguire G. Therapeutics from adult stem cells and the hype curve. ACS Med. Chem. Lett. 7(5), 441-443 (2016).
- 44. Maguire G. Stem cells, part of the innate and adaptive immune systems, as an antimicrobial for Covid-19. ACS Med. Chem. Lett. (2020) (In Press).
- Bernheim A, Mei X, Huang M et al. Chest CT findings in coronavirus disease-19 (COVID-19): relationship to duration of infection. Radiology https://doi.org/10.1148/radiol.2020200463 (2020) (Epub ahead of print).



- 46. Bioregenerative Sciences, Inc: US9545370B2 (2015).
- 47. Bioregenerative Sciences, Inc: US9446075B2 (2014).
- 48. Bioregenerative Sciences, Inc: US2014205563A1 (2014).
- 49. Bioregenerative Sciences, Inc: US2013302273A1 (2013).